

## REVIEW ARTICLE

# The Plant Kingdom as a Source of Anti-ulcer Remedies

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Phylogenetic agents have traditionally been used by herbalists and indigenous healers for the prevention and treatment of peptic ulcer. This article reviews the anti-acid/anti-peptic, gastro-protective and/or anti-ulcer properties of the most commonly employed herbal medicines and their identified active constituents. Botanical compounds with anti-ulcer activity include flavonoids (i.e. quercetin, naringin, silymarin, anthocyanosides, sophoradin derivatives) saponins (i.e. from *Panax japonicus* and *Kochia scoparia*), tannins (i.e. from *Linderae umbellatae*), gums and mucilages (i.e. gum guar and myrrh). Among herbal drugs, liquorice, aloe gel and capsicum (chilli) have been used extensively and their clinical efficacy documented. Also, ethnomedical systems employ several plant extracts for the treatment of peptic ulcer. Despite progress in conventional chemistry and pharmacology in producing effective drugs, the plant kingdom might provide a useful source of new anti-ulcer compounds for development as pharmaceutical entities or, alternatively, as simple dietary adjuncts to existing therapies. Copyright © 2000 John Wiley & Sons, Ltd.

*Keywords:* gastric ulcer; gastro-protective effect; flavonoids; saponins; tannins; gum and mucilages.

## INTRODUCTION

There is a balance in the stomach between the aggressive digestive capabilities of acid plus pepsin and the mucosal barrier. Ulceration occurs when there is a disturbance of the normal equilibrium caused by either enhanced aggression or diminished mucosal resistance. Several factors are implicated in the pathogenesis of gastric ulcer. These include increased acid-pepsin secretion, impaired bicarbonate neutralization, impaired mucus secretion and precipitate lesions on the mucosal layer (Kent Lloyd and Debas, 1994). Acid and pepsin secretion must be considered together because in practice it is difficult to distinguish the effects of each alone (Hersey, 1994). Drug treatment of peptic ulcers is targeted at either counter-acting aggressive factors (acid plus pepsin, active oxidants, PAF, leukotrienes, endothelins, bile or exogenous factors including NSAIDs) or stimulating the mucosal defences (mucus, bicarbonate, normal blood flow, prostaglandins, nitric oxide) (Tepperman and Jacobson, 1994). The ideal aims of treatment of peptic ulcer disease are to relieve pain, heal the ulcer and delay ulcer recurrence. To date, no drug meets all the goals of therapy.

In recent years, gastric ulcer has also been associated with infection of gastrointestinal mucosal tissue by *Helicobacter pylori* (Tepperman and Jacobson, 1994). About 70% of patients with peptic ulcer disease are infected by *Helicobacter pylori* and eradication of this

microorganism seems to be curative for this disease (Blaser, 1998).

This article reviews drugs derived from botanical sources more commonly used (or extensively studied) in the world for peptic ulcer and, if reported, the anti-ulcer mechanism postulated. While there are other actions on the stomach (e.g. emesis, motility) of the substances reviewed, this article will be concerned only with the anti-ulcer and gastro-protective effects.

## ACTIVE PRINCIPLES WITH ANTI-ULCER ACTIVITY

### Flavonoids

Flavonoids are a group of about 4000 naturally occurring compounds with a wide range of biological effects, including anti-ulcer activity. They are important constituents of the human diet (a daily diet contains approximately 1 g of flavonoids per day) and are also found in several medicinal plants used in folk medicine around the world (Di Carlo *et al.*, 1999). Several mechanisms have been proposed to explain the gastro-protective effect of flavonoids; these include increase of mucosal prostaglandin content (Alcaraz and Hoult, 1985), decrease of histamine secretion from mast cells by inhibition of histidine decarboxylase (Bronner and Landry, 1985) and inhibition of *Helicobacter pylori* growth (Beil *et al.*, 1995). In addition, flavonoids have been found to be free radical scavengers (Baumann *et al.*, 1980; Cavallini *et al.*, 1978; Salvayre *et al.*, 1982); free radicals play an important role in ulcerative and erosive

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**Table 1. Flavonoids with anti-ulcer activity**

Flavonoid	Ulcer model	Reference
Anthocyanosides	Pylorus-ligated, reserpine, phenylbutazone	Magistretti <i>et al.</i> , 1998; Cristoni and Magistretti, 1987
Catechin	Stress	Lorenz <i>et al.</i> , 1975
Genistin	Phenylbutazone, serotonin pylorus-ligated, stress, reserpine	Rainova <i>et al.</i> , 1988
Hypolaetin-8-glucoside	Stress, ethanol, acetylsalicylic acid	Alcaraz and Houtl, 1985; Villar <i>et al.</i> , 1984
Kaempferol	Ethanol	Izzo <i>et al.</i> , 1994
Leucocyanidin	Aspirin	Lewis <i>et al.</i> , 1999
Luteolin-7-glycoside,	Pylorus-ligated, stress, reserpine, phenylbutazone, serotonin	Rainova <i>et al.</i> , 1988
5-Methoxyflavone	Indomethacin	Blank <i>et al.</i> , 1997
Myricetin 3-O-D-galactoside	Stress, pylorus-ligated, ethanol	Reyes <i>et al.</i> , 1996
Naringin	Ethanol, stress, pylorus-ligated	Martin <i>et al.</i> , 1993; Motilva <i>et al.</i> , 1993
Quercetin	Stress, ethanol, reserpine	Izzo <i>et al.</i> , 1994; Di Carlo <i>et al.</i> , 1994; Martin <i>et al.</i> , 1993
Rutin	Ethanol	Izzo <i>et al.</i> , 1994
Silymarin	Stress	Alarcon de la Lastra, 1992
Ternatin	Ethanol, indomethacin, stress	Rao <i>et al.</i> , 1997
Vexibinol	HCl, ethanol	Yamahara <i>et al.</i> , 1990

lesions of the gastrointestinal tract. In relation to their low toxicity and to the properties reported, flavonoids could have a therapeutic potential ideal for treatment of gastrointestinal diseases associated with *Helicobacter pylori* infection, i.e. type B gastritis and duodenal ulcer (Di Carlo *et al.*, 1999).

Anti-ulcer flavonoids are shown in Table 1. The most studied are naringin, quercetin, silymarin, anthocyanosides and sophoradin derivatives. Moreover, several extracts containing flavonoids have been found to exert gastro-protective activity (Di Carlo *et al.*, 1999), although the precise chemical composition, in many cases, is unknown.

**Quercetin.** Quercetin is the most abundant of the flavonoid molecules and it is found in many medicinal botanicals, including *Thea sinensis*, *Glycyrrhiza glabra*, *Hypericum perforatum*, *Ginkgo biloba* and many others. It has been reported to prevent gastric mucosal lesions produced by cold-restraint stress and pylorus-ligation (Martin *et al.*, 1988; 1993), reserpine (Manicheva and Barnaulov, 1984) and ethanol or acidified ethanol (Alarcon de la Lastra *et al.*, 1992; Izzo *et al.*, 1994; Di Carlo *et al.*, 1994). Quercetin increases the amount of neutral glycoproteins in the gastric mucosa (Di Carlo *et al.*, 1999). As these proteins are the most abundant and possibly the most important in the gastric mucosa (Marti-Bonmati *et al.*, 1980), it can be assumed that their quantitative replacement is a return to normality of the mucosa, and thus a recovery of the defensive capacity against aggression from absolute ethanol.

Stimulation of local prostaglandin production is possibly one protective mechanism as quercetin stimulates the enzyme cyclooxygenase (Moroney *et al.*, 1988) and, in addition, the cyclooxygenase inhibitor, indomethacin, reverses the protective effect of quercetin on ethanol-induced ulceration (Robak and Gryglewski, 1988). The increase in prostaglandin synthesis may explain the increase in the amount of mucus observed and its participation in ulcer prevention. Other possible mechanisms include inhibition of the gastric proton pump (Di Carlo *et al.*, 1999), inhibition of the lipoxigenase

pathway (Moroney *et al.*, 1988), inhibition of platelet activating factor synthesis (Izzo *et al.*, 1994), inhibition of lipid peroxidation (Alarcon de la Lastra *et al.*, 1994) and scavenging of free radicals associated with a significant enhancement in glutathione peroxidase activity (Martin *et al.*, 1998). An interesting aspect of the anti-ulcer effect of quercetin is that it has been shown to inhibit the growth of *Helicobacter pylori* in a dose-dependent manner *in vitro* (Beil *et al.*, 1995).

**Naringin.** Naringin has been shown to prevent gastric mucosal ulceration in several animal models, including restraint stress, pyloric occlusion and ethanol-induced chronic ulcer (Parmar, 1983; Martin *et al.*, 1993; Motilva *et al.*, 1992; 1993).

In ethanol-treated rats, naringin significantly reduced the ulcer index and increased the hexosamine content of the gastric mucus without affecting prostaglandin E<sub>2</sub> and the total protein content (Martin *et al.*, 1994). Thus, the gastro-protective action of naringin could be explained, at least in part, through a complex non-prostaglandin-dependent mechanism that involves an increase in the glycoprotein content and viscosity of the gastric mucosa. Naringin also possesses antioxidant and superoxide anion scavenger properties (Robak *et al.*, 1988; Martin *et al.*, 1994) which could contribute to its gastro-protective effect.

**Silymarin.** Silymarin is a flavanolignan complex which can be extracted from the fruit (*Cardui marie fructus*) of the milk thistle, *Silybum marianum* (Samuelsson, 1999).

Silymarin has been found effective in some types of experimentally induced gastric ulcers. Treatment with silymarin prevents, in a highly significant way, the ulceration induced by cold-restraint stress and histamine output in pylorus-ligated rats but does not prevent the formation of ethanol-induced gastric lesions (Alarcon de la Lastra *et al.*, 1992). The anti-ulcerogenic effect of silymarin could be related to its inhibitory mechanism on the lipoxigenase pathway, avoiding leukotriene synthesis (Bindoli *et al.*, 1977; Alarcon de la Lastra *et al.*, 1992).

**Table 2. Plants containing saponins with anti-ulcer activity**

Botanical name	Part plant	Ulcer model	Reference
<i>Calendula officinalis</i>	Rhizome	Caffeine-arsenic, butadione, pylorus-ligated	latsyno <i>et al.</i> , 1978
<i>Calliandra portoticensis</i>	Leaves	Stress, pylorus-ligated, <i>E. coli</i>	Aguwa and Lawal, 1988
<i>Glycyrrhiza glabra</i>	Root	Acetic acid, pylorus ligated	Ottenjann and Rosh, 1970
<i>Kochia scoparia</i>	Fruit	Ethanol, indomethacin	Matsuda <i>et al.</i> , 1998
<i>Panax binnatifidus</i>	Rhizome	Psychological stress	Nguyen <i>et al.</i> , 1996; Houg <i>et al.</i> , 1998
<i>Panax japonicus</i>	Rhizome	Stress, HCl	Yamahara <i>et al.</i> , 1987
<i>Pyrenacantha staudtii</i>	Leaves	Indomethacin, serotonin, stress	Aguwa and Okunji, 1986
<i>Rhigiocarya racemifera</i>	Leaves	Indomethacin, reserpine, serotonin	Aguwa, 1985; 1986
<i>Spartium junceum</i>	Flowers	Ethanol	Yesilada and Takaishi, 1999
<i>Veronica officinalis</i>	Aerial parts	Indomethacin, reserpine	Scarlat <i>et al.</i> , 1985

**Anthocyanosides.** Anthocyanosides, extracts from *Vaccinium myrtillus*, exert a significant preventive and curative anti-ulcer activity. Protective effects have been observed not only on ulcer induced by ligation of the pylorus, by reserpine and by phenylbutazone, in which oral treatment was given before or simultaneously with ulcer formation, but also in ulcer induced by restraint or by acetic acid, in which the animal received treatment when the lesions were already established (Magistretti *et al.*, 1988). The anti-ulcer activity is not exerted through a blockade of gastric secretion but it is, in part, attributed to the increase in the mucus of the stomach wall. Anthocyanosides are thought to act by influencing the biosynthesis of the mucopolysaccharides, thus improving the efficiency of the mucus barrier at the gastric level (Cristoni and Magistretti, 1987; Magistretti *et al.*, 1988).

**Sophoradin derivatives.** Sophoradin has been isolated from the root of the ancient Chinese medical plant *Sophora subprostrata*, a plant used in China for the treatment of digestive diseases (Kyogoku *et al.*, 1979). Flavonoids derived from sophoradin are known to exhibit gastro-protective and ulcer healing properties. Solon, a synthetic flavonoid derived from sophoradin, has been shown to have anti-ulcer and gastro-protective effects by influencing the formation and metabolism of prostaglandins in the gastric mucosa (Konturek *et al.*, 1986).

Recently, Brzozowski *et al.* (1998) have reported that SU-840, a novel synthetic flavonoid derivative of sophoradin reduced gastric acid and pepsin secretion and gastric lesions induced by ethanol, acidified aspirin or restraint stress, an effect due in part to nitric oxide and prostaglandin production.

## Saponins

Saponins are widely distributed in plants and are a particular form of glycosides. They are so-called because of their soap-like effect, which is due to their surfactant properties. They also have haemolytic properties and, when injected into the blood stream, are highly toxic. When taken by mouth saponins are comparatively harmless. According to the structure of the aglycone or sapogenin two kinds of saponin are recognized, the steroidal and triterpenoid type (Samuelsson, 1999).

Plant materials often contain triterpenoid saponins in considerable amounts. With regard to plants with anti-ulcer activity, liquorice root contains about 2%–12% of

glycyrrhizic acid and the seeds of the horse-chestnut up to 13% of aescin (Newall *et al.*, 1996). Several plants containing high amounts of saponins have been shown to possess anti-ulcer activity in several experimental ulcer models (Table 2). Among these, saponins isolated from the rhizome of *Panax japonicus* and the fruit of *Kochia scoparia* (which contain approximately 20% of saponins) have been demonstrated to possess gastro-protective properties (Matsuda *et al.*, 1998). The same authors reported that some oleanolic acid oligoglycosides, extracted from *P. japonicus* and *K. scoparia*, showed protective effects on ethanol- and indomethacin-induced gastric damage. Moreover, a methanol extract of *P. japonicus* rhizome was demonstrated to possess protective activity also on stress- or HCl-induced ulcers (Yamahara *et al.*, 1987). The protective activities of all these active saponins are not due to inhibition of gastric acid secretion but probably due to activation of mucous membrane protective factors (Saito *et al.*, 1977).

Aescin is a mixture of saponins obtained from the seeds of *Aesculus hippocastanum* (horse-chestnut). Aescin has been shown to possess anti-ulcer activity in various ulcer models (cold restraint and pylorus-ligated) (Marhuenda *et al.*, 1993), an effect which is, in part, due to inhibition of gastric acid and pepsinogen secretion. However, aescin also prevents gastric lesions due to absolute ethanol (Marhuenda *et al.*, 1994), a model of gastric ulceration in which acid and pepsin do not play a significant role; thus, the protective effect of aescin involves other mechanisms, such as improvement of blood flow (Marhuenda *et al.*, 1994). It is unlikely that prostaglandins or the maintenance of surface mucus gel could play a role in aescin-induced gastro-protection as this saponin does not enhance prostaglandin E<sub>2</sub> levels nor does it increase the amount (or the composition) of mucus in ethanol-induced gastric ulceration (Marhuenda *et al.*, 1994).

## Tannins

Tannins are used in medicine primarily because of their astringent properties, which are due to the fact that they react with the proteins of the layers of tissue with which they come into contact (Samuelsson, 1999). Tannins are known to 'tan' the outermost layer of the mucosa and to render it less permeable and more resistant to chemical and mechanical injury or irritation (Asuzu and Onu, 1990); however, the correlation between the molecular

**Table 3. Plants containing tannins with anti-ulcer activity**

Botanical name	Part plant	Ulcer model	Reference
<i>Calliandra portoticensis</i>	Leaves	Stress, pylorus-ligated, <i>E. coli</i>	Aguwa and Lawal, 1988
<i>Entandrophragma utile</i>	Bark	Ethanol	John and Onabanjo, 1990
<i>Linderae umbellatae</i>	Stem	Stress	Ezaki <i>et al.</i> , 1985
<i>Mallotus japonicus</i>	Bark	(Clinical study)	Saijo <i>et al.</i> , 1989
<i>Rhigiocarya racemifera</i>	Leaves	Indomethacin, reserpine, serotonin	Aguwa, 1985; 1986
<i>Veronica officinalis</i>	Aerial parts	Indomethacin, reserpine	Scarlat <i>et al.</i> , 1985

structures of tannins and the astringent/anti-ulcer activity is not known.

When a low concentration of tannins is applied to the mucosa, only the outermost layer is tanned, becoming less permeable and affording an increased protection to the subjacent layers against the action of bacteria, chemical irritation, and, to a certain extent, against mechanical irritation. High concentrations of tannins cause coagulation of the proteins of the deeper layer of the mucosa, resulting in inflammation, diarrhoea and vomiting.

Several plants with anti-ulcer activity which contain tannins have been reported (Table 3). A crude extract of *Linderae umbellatae* exhibited anti-peptic and anti-ulcerogenic activity, and these effects were considered ascribable to the presence of tannins or related compounds (Ezaki *et al.*, 1985). Nine condensed tannins (monomers, dimers, trimers and tetramers) have been isolated and their anti-peptic and anti-ulcer activity confirmed experimentally (pylorus-ligated in rats and stress-induced gastric lesions in mice) (Ezaki *et al.*, 1985). Marked differences were observed among monomers, dimers, trimers and tetramers. Monomers and dimers, did not inhibit peptic activity *in vitro*, while trimers displayed higher inhibition of peptic activity than tetramers (though tetramers showed higher astringency than trimers). In pylorus-ligated mice, trimers and tetramers clearly suppressed the peptic activity of gastric juice and also monomers and dimers slightly suppressed the peptic activity of mouse gastric juice *in vivo* (Ezaki *et al.*, 1985). As monomers and dimers are inactive *in vitro* it is possible that their activity *in vivo* is not related to the direct inhibition of pepsin, but related to influence on the secretion mechanism of pepsin.

### Gums and mucilages

Gums and mucilages are usually brittle, amorphous, transparent or translucent substances, which readily absorb water to form gelatinous masses or viscous colloidal solutions. The colloidal character of gums, mucilages and other mucoids accounts largely for their use as therapeutic agents. Mucilaginous drugs have the property of covering and protecting the mucosa of the stomach and are used in the treatment of gastric ulcer. Plants containing mucilages traditionally used in several countries in the treatment of gastric ulcer include *Althaea officinalis*, *Cetraria islandica*, *Malva sylvestris*, *Matricaria chamomilla* and *Aloe* species (Capasso and Grandolini, 1999)

Guar gum is obtained from the endosperm of the seed of *Cyamopsis tetragonolobus*, a plant long cultivated in

India and Pakistan and nowadays also grown in the United States (Samuelsson, 1999). The main constituent is a galactomannan, a  $\beta$ -1,4-linked D-mannose linear polysaccharide with an  $\alpha$ -1,6-linked D-galactose residue attached to every other D-mannose unit. Gum guar increased the healing rate of stress-induced gastric ulcers in the rat. Proposed mechanisms are reduced acidity, increased local mucosal supply of energy and mechanical protection (Harju and Sajanti, 1987; 1991). Moreover, guar gum, in duodenal ulcer patients, decreases gastric acidity and the rate of emptying of gastric contents, probably because of its effects on viscosity and neutralization of gastric acidity (Harju and Larimi, 1985; Harju, 1984; 1985).

Myrrh, an oleo-gum-resin obtained from *Commiphora molmol*, contains up to 60% gum and up to 40% resin (Newall *et al.*, 1996). Myrrh pre-treatment produced a dose-dependent protection against the ulcerogenic effects of different necrotizing agents such as ethanol, indomethacin, sodium hydroxide or hypertonic sodium chloride (Al-Harbi *et al.*, 1997). The protective effect of myrrh is attributed to its effect on mucus production or increase in nucleic acid and non-protein sulphhydryl concentration, which appears to be mediated through its free-radical scavenging, thyroid-stimulating and prostaglandin-inducing properties.

## HERBAL DRUGS WITH ANTI-ULCER ACTIVITY

### Liquorice

Liquorice, the root and rhizome of different varieties of *Glycyrrhiza glabra* (Leguminosae), has been extensively used in medicine for its anti-ulcer activity (Ottenjann and Rosch, 1970). The principal constituent of liquorice is glycyrrhizic acid, a triterpenoid saponin. This chemical class is known to offer protection against ulcers. Takagi *et al.* (1969) showed that glycyrrhizin (the potassium or calcium salt of glycyrrhizic acid) inhibited ulcer in Shay rats and also prevented experimental gastric ulcers caused by acetic acid administration. To avoid the side-effects of glycyrrhizin in liquorice preparations, a glycyrrhizin-free fraction of liquorice extracts was studied (Takagi and Ishii, 1967). The glycyrrhizin-free fraction, called FM 100 fraction, was found to be effective for gastric ulcers in rats by inhibiting gastric juice secretion. The protective effect of FM 100 fraction has been confirmed by clinical observation (Takagi *et al.*, 1969). Saitoh *et al.* (1976) isolated new isoflavonoids and chalcones from the fraction FM 100, which could

contribute to the anti-ulcer activity of liquorice. Simple derivatives of glycyrrhetic acid (the genin of glycyrrhizinic acid) such as carbenoxolone (an ester derivative) and deglycyrrhizinized liquorice (DGL), have been used extensively in gastric ulcer treatment (Habib *et al.*, 1979; Bennett *et al.*, 1980).

Carbenoxolone derives from the hydrolysis of glycyrrhizinic acid after its extraction from liquorice root. Carbenoxolone was the first drug found to accelerate peptic ulcer healing by a mechanism not involving the inhibition of acid secretion (Baron, 1977). Several mechanisms of action have been proposed to explain the pharmacological activity of carbenoxolone. Thus, carbenoxolone stimulates gastric mucus production (Bickel and Kauffman, 1981), enhances the rate of incorporation of various sugars into gastric mucosal glycoproteins (Johnston *et al.*, 1975), promotes mucosal cell proliferation (Van Huis and Kramer, 1981), inhibits mucosal cell exfoliation (Domschke *et al.*, 1977), inhibits prostaglandin degradation (Peskar, 1980), increases the release of PGE<sub>2</sub> and reduces the formation of thromboxane B<sub>2</sub> (Aguwa and Okunji, 1986) and regulates DNA and protein synthesis rates in gastric mucosa (Van Huis and Kramer, 1981). More recently nitric oxide has been claimed to contribute to the anti-ulcer effect of carbenoxolone (Peskar *et al.*, 1991; Franco *et al.*, 1993). In view of the wide range of alternative therapies now available and of the numerous side effects, carbenoxolone use has been limited; indeed carbenoxolone produces effects similar to those of aldosterone excess (sodium retention and hypokalaemia leading to hypertension, oedema and cardiac failure) (Newall *et al.*, 1996).

DGL is an effective anti-ulcer agent and, unlike carbenoxolone, it does not have fluid retaining properties. DGL protected against gastric ulceration in rats induced by pyloric ligation and increased healing of peptic ulcer in patients (Rees *et al.*, 1979). Moreover, DGL reduced bile acid-induced hydrogen ion back-diffusion across the canine gastric mucosa and diminished acute gastric mucosal damage due to aspirin or aspirin plus taurodeoxycholic acid (Morgan *et al.*, 1983).

### Aloe gel

Two distinct preparations of *Aloe* plants are most used medicinally. The resinous leaf exudate (named aloe) is used as a laxative, and the mucilaginous gel (named aloe vera or aloe gel) from the leaf parenchyma is used as a remedy against a variety of skin disorders (Reynolds and Dweck, 1999). Aloe gel had a prophylactic effect and was also curative if given as a treatment for stress-induced gastric ulceration in rats (Galal *et al.*, 1975; Parmar *et al.*, 1986). A lectin fraction (glycoprotein) from *Aloe arborescens*, Aloctin A, had anti-ulcer effect in rats (Saito *et al.*, 1989), while another high molecular weight fraction, not containing glycoprotein, was very effective in healing ulcer induced by mechanical or chemical stimuli but not those induced by stress (Teradaira *et al.*, 1993). This fraction contained substances with molecular weights between 5000 and 50000 Da, which were considered to both suppress peptic ulcers and to heal chronic gastric ulcers. In addition, a component from Cape Aloe exudate, named aloe ulcin, suppressed ulcer growth and L-histidine decarboxylase in rats (Yamamoto, 1973). An early clinical study found that oral adminis-

tration of aloe gel was effective in the treatment of peptic ulcer (Blitz *et al.*, 1963). By contrast, in a more recent study, it has been shown that a health product containing aloe vera gel did not prevent lesion formation associated with cold-restraint or ethanol administration in animals (Koo, 1994).

### Capsicum (chilli)

Capsicum, also known as chilli or paprika, is the fruit of various *Capsicum* species (Samuelsson, 1999). It is widely used as a spice and, traditionally, it has been used internally for colic, flatulent dyspepsia, chronic laryngitis, insufficiency of peripheral circulation and externally for neuralgia (Newall *et al.*, 1996). Capsaicin (the active pungent ingredient) has been used extensively as a probe to elucidate the function of sensory neurons in various organs and systems (including the stomach), because of its ability to excite and later defunctionalize a subset of primary afferent neurons (Abdel-Salam *et al.*, 1997b).

Chilli, as well as capsaicin, has a protective effect on ethanol- or aspirin-induced lesion formation in the rat gastric mucosa (Kang *et al.*, 1995a); in addition capsaicin and long-term chilli intake (360 mg daily for 4 weeks) protects against haemorrhagic shock-induced gastric mucosal injury in the rat, an effect which may be mediated by capsaicin-sensitive afferent neurons (Teng *et al.*, 1998). The pathophysiological basis of gastric protection by chilli (capsaicin) remains unclear. An increase in gastric mucosal blood flow has been described and this may be related to the release of calcitonin gene-related peptide and nitric oxide rather than to the production of prostaglandins (Lambrecht *et al.*, 1993). The protective effect of capsaicin could also involve vanilloid receptors because resiniferatoxin, an ultrapotent analogue of capsaicin present in the latex of *Euphorbia resinifera* (Euphorbium), also displays anti-ulcer activity (Abdel Salam *et al.*, 1997a) and both capsaicin and resiniferatoxin act on vanilloid (capsaicin) receptors (Szallasi and Blumberg, 1999).

Chilli causes dyspepsia in patients with and without ulcer, and ulcer patients are often advised to avoid its use (Kang *et al.*, 1992). Therefore, the peptic ulcer patients consume less chilli than controls (Kang *et al.*, 1995b). Nevertheless, epidemiological and clinical data suggest that chilli ingestion may have a beneficial effect on human peptic ulcer disease. For examples chilli has a gastro-protective effect on aspirin-induced gastric mucosal injury in humans (Yeoh *et al.*, 1995). In addition, Kumar *et al.* (1984) showed that chilli ingestion has no detrimental effect on the healing rates of duodenal ulcer patients on treatment and it does not cause macroscopic gastroduodenal mucosal injury in humans (Kang *et al.*, 1988). By contrast, others have shown that chilli increased DNA loss and microbleeding (Desai *et al.*, 1972; Myers *et al.*, 1987) suggesting the occurrence of cellular damage.

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## HERBAL FORMULATIONS

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There are many plants used in anti-ulcer herbal formulations that have not yet been individually investigated for

**Table 4. Plants with anti-ulcer activity from ethnopharmacological studies**

Botanical name	Part plant	Extract type	Ulcer model	Reference
<i>Alpinia galanga</i>	Rhizome	Ethanol	Stress, pylorus-ligated, ethanol, HCl	Al-Yahya <i>et al.</i> , 1990
<i>Amphipterygium adstringens</i>	Stem bark	Methanol	Ethanol	Navarrete <i>et al.</i> , 1998
<i>Anchusa strigosa</i>	Root	Aqueous	Ethanol	Disi <i>et al.</i> , 1998
<i>Artemisia herba-alba</i>	Leaves	Aqueous	Ethanol	Gharzouli <i>et al.</i> , 1999
<i>Astronium urundeuva</i>	Bark	Aqueous	Aspirin, stress, histamine	Rao <i>et al.</i> , 1987
<i>Atractylodis lancea</i>	Rhizome	Acetone	Ethanol, HCl	Matsuda <i>et al.</i> , 1998
<i>Azadirachta indica</i>	Leaves	Aqueous	Stress, ethanol	Garg <i>et al.</i> , 1993
<i>Baccharis triptera</i>	Small branches	Aqueous	Pylorus-ligated, stress, indomethacin	Gamberini <i>et al.</i> , 1991
<i>Bauhinia racemosa</i>	Flower buds	Methanol	Aspirin	Akhtar and Ahmad, 1995
<i>Bryophyllum pinnatum</i>	Leaves	Methanol	Aspirin, indomethacin, serotonin, reserpine, stress, ethanol, pylorus-ligated, acetic acid	Pal and Nag Chaudhuri, 1991
<i>Caesalpinia ferrea</i>	Stem	Crude	Acetic acid	Bacchi <i>et al.</i> , 1995
<i>Camellia sinensis</i>	Leaves	Aqueous	Stress, ethanol, aspirin, indomethacin, reserpine, histamine, serotonin	Maity <i>et al.</i> , 1995
<i>Cassia nigrans</i>	Leaves	Ethanol	Aspirin, pylorus-ligated	Maity <i>et al.</i> , 1995; Akah <i>et al.</i> , 1998
<i>Cistus incanus</i>	Aerial part	Aqueous	HCl, ethanol, reserpine, serotonin	Attaguile <i>et al.</i> , 1995
<i>Curcuma longa</i>	Rhizome	Ethanol	Pylorus-ligated, cold-restraint stress, indomethacin, reserpine, ethanol, HCl	Rafatullah <i>et al.</i> , 1990
<i>Diodia sarmentosa</i>	Whole plant	Ethanol	Aspirin, pylorus-ligated	Akah <i>et al.</i> , 1998
<i>Entandrophragma utile</i>	Bark	Aqueous	Ethanol	John and Onabanjo, 1990
<i>Eremomastax speciosa</i>	Leaves	Aqueous	Ethanol, HCl, pylorus-ligated	Tan <i>et al.</i> , 1996
<i>Ficus exasperata</i>	Leaves	Ethanol	Aspirin, pylorus-ligated	Akah <i>et al.</i> , 1998
<i>Laurus nobilis</i>	Seeds	Aqueous, oil	Ethanol	Afifi <i>et al.</i> , 1997
<i>Maytenus aquifolium, ilicifolia</i>	Leaves	Aqueous	Indomethacin, cold-restraint stress	Souza-Formigoni <i>et al.</i> , 1991
<i>Microgramma squamulosa</i>	Rhizome	Crude, ethanol, water	Stress, ethanol, HCl, acetic acid	Suffredini <i>et al.</i> , 1999
<i>Mikania cordata</i>	Root	Methanol	Stress, ethanol, aspirin, phenylbutazone, pylorus-ligated	Bishayee and Chatterjee, 1994
<i>Moringa pterygosperma</i>	Flower buds	Methanol	Aspirin	Akhtar and Ahmad, 1995
<i>Pistacia lentiscus</i>	Resin from stem		Pylorus-ligated, aspirin, phenylbutazone Reserpine-restraint plus cold stress, ethanol	Al-Said <i>et al.</i> , 1986
<i>Pluchea indica</i>	Root	Methanol	Indomethacin, ethanol, aspirin, stress, reserpine	Sen <i>et al.</i> , 1992
<i>Punica granatum</i>	Fruit peel	Aqueous	Ethanol	Gharzouli <i>et al.</i> , 1999
<i>Pyrenacantha staudtii</i>	Leaves	Aqueous	Aspirin, indomethacin, serotonin, reserpine	Aguwa and Mittal, 1981
<i>Quercus ilex</i>	Root bark	Aqueous	Ethanol	Gharzouli <i>et al.</i> , 1999
<i>Saussurea lappa</i>	Root	Acetone	Stress	Yamahara <i>et al.</i> , 1985
<i>Stachytarpheta cayennensis</i>	Whole plant	Aqueous	Stress, ethanol, pylorus-ligated	Vela <i>et al.</i> , 1997
<i>Stryphnodendron adstringens</i>	Aerial parts	Total extract	Stress, ethanol, indomethacin	Audi <i>et al.</i> , 1999
<i>Styrax camporum</i>	Stem	Ethyl acetate	Acetic acid	Bacchi <i>et al.</i> , 1995
<i>Swertia chirata</i>	Whole plant	Ethanol	Indomethacin, pylorus-ligated, ethanol	Rafatullah <i>et al.</i> , 1993
<i>Synclisia scabrida</i>	Leaves	Ethanol	Aspirin, pylorus-ligated	Akah <i>et al.</i> , 1998
<i>Tanacetum vulgare</i>	Aerial parts	Chloroform	Ethanol	Tournier <i>et al.</i> , 1999
<i>Trianthema pentandra</i>	Whole plant	Methanol	Aspirin	Akhtar and Ahmad, 1995
<i>Trichosanthes kirilowii</i>	Fruit	Ethanol	Stress, histamine, serotonin, ethanol, HCl, pylorus-ligated	Takano <i>et al.</i> , 1990
<i>Vernonia kotschyana</i>	Root	Aqueous	Stress, indomethacin, ethanol, pylorus-ligated	Sanogo <i>et al.</i> , 1996; Germanó <i>et al.</i> , 1996
<i>Zingiber officinalis</i>	Root	Methanol, acetone	HCl/ethanol	Yamahara <i>et al.</i> , 1988

their efficacy in this condition. Mitra *et al.*, (1996) reported the anti-ulcer effect of a herbal formulation, UL-409, consisting of six medicinal plants, namely *Glycyrrhiza glabra* L. (Papilionaceae; Root), *Saussurea lappa* C.B. Clarke (Compositae; Rott), *Aegle marmelos* Corr. (Rutaceae; Fruit), *Foeniculum vulgare* Mill. (Umbelliferae; Seed), *Rosa damascena* Mill. (Rosaceae; Flower Petals) and *Santalum album* L. (Santalaceae; Stem). This herbal formulation was found to increase the stomach mucus and decrease the acid volume, free and total acid content in rats. Moreover this formulation significantly prevented the occurrence of stress-induced ulceration, and significantly inhibited gastric ulceration induced by ethanol, aspirin, pylorus-ligation, histamine and indomethacin in rats and guinea-pigs (Mitra *et al.*, 1996; Kulkarni and Goel, 1996). On the basis of these observations, it was concluded that this formulation possesses anti-ulcer activity due to a modulation of defensive factors so improving gastric cytoprotection.

Manonmani and co-workers (1994) reported the anti-ulcerogenic effect of an Ayurvedic formulation, Cauvery-100 (C-100). C-100 consists of dasamoolam (a preparation consisting of 10 medicinal plants) plus 13 other medicinal plants, namely *Emblia officinalis* L. (Euphorbiaceae, fruit), *Cuminum cyminum* L. (Umbelliferae, fruit), *Trema orientalis* L. Blume (Urticaceae, root), *Vitis vinifera* L. (Vitaceae, fruit), *Carum carvi* L. (Umbelliferae, fruit), *Withania somnifera* L. Dunal (Solanaceae, root), *Eclipta alba* L. Hassk (Compositae, whole plant), *Embelia ribes* Burm (Myrsinaceae, fruit), *Citrum limon* L. Burm (Rutaceae, fruit rind), *Picrorrhiza kurroa* Benth (Scrophulariaceae, root), *Glycyrrhiza glabra* L. (Papillonace, weakstem), *Aegle marmelos* L. Corr. Serr (Rutaceae, root) and *Zingiber officinale* Roscoe (Zingiberaceae, rhizome). This formulation was found to decrease the number of lesions induced by indomethacin and to decrease the volume and total acidity of gastric juice in rats. It was also found that C-100 increased the levels of hexose, hexosamine and sialic acid (a substance normally decreased in the untreated ulcer group), while decreasing the protein level (normally raised in the untreated ulcer group). The authors concluded that C-100 could act both to decrease acidity and to increase mucosal defence in the gastric region.

## ETHNOPHARMACOLOGICAL STUDIES

Gastrointestinal disorders are one of the most important causes of morbidity for the populations of non-industrialized countries. Many pharmaceutical products employed for the treatment of gastroduodenal ulcers and peptic diseases, have decreased the morbidity rates, but they can produce many adverse effects. Moreover, these pharmaceutical products are too expensive for poorer populations. The importance of ethnopharmacological studies is emphasized by the observation that the first

drug effective against gastric ulcer was carbenoxolone, discovered as a result of research on a commonly used indigenous plant, *Glycyrrhiza glabra*. Also, studies on cabbage, previously employed as an anti-ulcer agent in folk medicine, has led to the development of gefarnate (Hossenbocus and Colin-Jones, 1974; Katoh *et al.*, 1998). Thus, a search among medicinal plants is still important, despite the progress in conventional chemistry and pharmacology in producing effective drugs. Countless plants known to indigenous medicine are used for the treatment of peptic ulcers; a rather conservative list is reported in Table 4. Among these, of particular interest are *Pteleopsis suberosa* and *Ocimum sanctum*.

The bark of *Pteleopsis suberosa* is commonly used in Mali for the treatment of gastric ulcers and was also claimed to be effective in a clinical trial (Mariko, 1989). Experimental studies have shown that a *Pteleopsis suberosa* decoction obtained from the bark of plants showed a protective action against ethanol- and indomethacin-induced gastric mucosa damage, an effect which is due in part to the stimulation of prostaglandin synthesis. A saponin fraction is most likely responsible for the biological activity (De Pasquale *et al.*, 1995; Germanò *et al.*, 1998).

*Ocimum sanctum* is commonly known as 'Tulsi' in Hindi and 'Holy basil' in English. The fixed oil of *Ocimum sanctum* has been shown to possess anti-ulcer activity against aspirin-, indomethacin-, alcohol-, histamine-, reserpine-, serotonin- and stress-induced ulceration in experimental animal models (Mandal *et al.*, 1993; Singh and Majumdar, 1999). Significant inhibition was also observed in gastric secretion and aspirin-induced gastric ulceration in pylorus-ligated rats. The anti-ulcer activity of the oil could be due to inhibition of the 5-lipoxygenase pathway or to leukotriene or histamine antagonistic properties (Singh and Majumdar, 1999).

## CONCLUSIONS

A variety of botanical products have been reported to possess anti-ulcer activity (especially from ethnopharmacological studies) but the documented literature has centred primarily on pharmacological action in experimental animals. Except for a few phytochemical compounds (i.e. aloe, liquorice and chilli), limited clinical data are available to support the use of herbs as gastroprotective agents and thus, the data on efficacy and safety are limited. Despite this, there are several botanical products with potential therapeutic applications because of their high efficacy and low toxicity. Finally, it should be noted that substances such as flavonoids, aescin, aloe gel and many others, that possess both anti-inflammatory and anti-ulcer activity are of particular therapeutic importance as most of the anti-inflammatory drugs used in modern medicine are ulcerogenic.

## REFERENCES

- Abdel Salam OM, Szolcsanyi J, Mozsik G. 1997a. The indomethacin-induced gastric mucosal damage in rats. Effect of gastric acid, acid inhibition, capsaicin-type agents and prostacyclin. *J Physiol Paris* **91**: 7–19.
- Abdel Salam OM, Szolcsanyi J, Mozsik G. 1997b. Capsaicin and the stomach. A review of experimental and clinical data. *J Physiol Paris* **91**: 151–171.
- Afifi FU, Khalil E, Tamimi SO, Disi A. 1997. Evaluation of the

- gastroprotective effect of *Laurus nobilis* seeds on ethanol induced gastric ulcer in rats. *J Ethnopharmacol* **58**: 9–14.
- Aguwa CN. 1985. Gastrointestinal effects of the extracts of *Rhigiocarya racemifera* (Menispermaceae). *Gen Pharmacol* **16**: 387–390.
- Aguwa CN. 1986. Pharmacologic effects of an aqueous extract of *Rhigiocarya racemifera*. *J Ethnopharmacol* **15**: 145–151.
- Aguwa CN, Lawal AM. 1988. Pharmacologic studies on the active principles of *Calliandra portoticensis* leaf extracts. *J Ethnopharmacol* **22**: 63–71.
- Aguwa CN, Mittal GC. 1981. Study of antiulcer activity of aqueous extract of leaves of *Pyrenacantha staudtii* (family Icacinaceae) using various models of experimental gastric ulcer in rats. *Eur J Pharmacol* **74**: 215–219.
- Aguwa CN, Okunji CO. 1986. Gastrointestinal studies of *Pyrenacantha staudtii* leaf extracts. *J Ethnopharmacol* **15**: 45–55.
- Akah PA, Orisakwe OE, Gamaniel KS, Shittu A. 1998. Evaluation of Nigerian traditional medicines: II. Effects of some Nigerian folk remedies on peptic ulcer. *J Ethnopharmacol* **62**: 123–127.
- Akhtar AH, Ahmad KU. 1995. Anti-ulcerogenic evaluation of the methanolic extracts of some indigenous medicinal plants of Pakistan in aspirin-ulcerated rats. *J Ethnopharmacol* **46**: 1–6.
- Alarcon de la Lastra C, Martin MJ, Marhuenda E. 1992. Gastric anti-ulcer activity of silymarin, a lipoxigenase inhibitor, in rats. *J Pharm Pharmacol* **44**: 929–931.
- Alarcon de la Lastra C, Martin MJ, Motilva V. 1994. Antiulcer and gastroprotective effects of quercetin: a gross and histologic study. *Pharmacology* **48**: 56–62.
- Alcaraz MJ, Hoult JR. 1985. Actions of flavonoids and the novel anti-inflammatory flavone, hypolaetin-8-glucoside, on prostaglandin biosynthesis and inactivation. *Biochem Pharmacol* **34**: 2477–2482.
- Al-Harbi MM, Quereshi S, Raza M, Ahmed MM, Afzal M, Shah AH. 1997. Gastric antiulcer and cytoprotective effect of *Commiphora molmol* in rats. *J Ethnopharmacol* **55**: 141–150.
- Al-Said MS, Ageel AM, Parmar NS, Tariq M. 1986. Evaluation of mastic, a crude-drug obtained from *Pistacia lentiscus* for gastric and duodenal anti-ulcer activity. *J Ethnopharmacol* **15**: 271–278.
- Al-Yahya MA, Rafatullah S, Mossa JS, Ageel AM, Al-Said MS, Tariq M. 1990. Gastric antisecretory, antiulcer and cytoprotective properties of ethanolic extract of *Alpinia galanga* Willd in rats. *Phytother Res* **4**: 112–114.
- Asuzu IU, Onu OU. 1990. Anti-ulcer activity of the ethanolic extract of *Combretum dolichopetalum* root. *Int J Crude Drug Res* **28**: 27–32.
- Attaguile G, Caruso A, Pennisi G, Savoca F. 1995. Gastroprotective effect of aqueous extract of *Cistus incanus* L. in rats. *Pharmacol Res* **31**: 29–32.
- Audi EA, Toledo DP, Peres PG *et al.*, 1999. Gastric antiulcerogenic affects of *Stryphnodendron adstringens* in rats. *Phytother Res* **13**: 264–266.
- Bacchi EM, Sertie JA, Villa N, Kanz H. 1995. Antiulcer action and toxicity of *Styrax camporum* and *Caesalpinia ferrea*. *Planta Med* **61**: 204–207.
- Baron JH. 1977. Effect of carbenoxolone sodium on human gastric acid secretion. *Gut* **18**: 721–722.
- Baumann J, Wurm G, Bruchhausen F. 1980. Prostaglandin synthetase inhibition by flavonoids and phenylic compounds in relation to their oxygen-scavenging properties. *Arch Pharm Weinheim* **313**: 330.
- Beil W, Birkholz C, Sewing KF. 1995. Effects of flavonoids on parietal cell acid-secretion, gastric mucosal prostaglandin production and *Helicobacter pylori* growth. *Arzneim Forsch* **45**: 697–700.
- Bennett A, Clark-Wibberley T, Stamford IF, Wright JE. 1980. Aspirin-induced gastric mucosal damage in rats: cimetidine and deglycyrrhizinated liquorice together give greater protection than low doses of either drug alone. *J Pharm Pharmacol* **32**: 151.
- Bickel M, Kauffman GL. 1981. Gastric gel mucus thickness: effect of distention, 16,16-dimethyl-prostaglandin E<sub>2</sub> and carbenoxolone. *Gastroenterology* **80**: 770–775.
- Bindoli A, Cavallini S, Siliprandi N. 1977. Accion inhibidora de la silimarina sobre la peroxidacion lipidica en los microsomas y mitocondrias hepaticos de la rata. *Biochem Pharmacol* **26**: 2405–2409.
- Bishayee A, Chatterjee M. 1994. Protective effects of *Mikania cordata* root extract against physical and chemical factor-induced gastric erosions in experimental animals. *Planta Med* **60**: 110–113.
- Blank MA, Ems BL, O'Brien LM *et al.* 1997. Flavonoid-induced gastroprotection in rats: role of blood flow and leukocyte adherence. *Digestion* **58**: 147–154.
- Blaser MJ. 1998. *Helicobacters* are indigenous to the human stomach: duodenal ulceration is due to changes in gastric microecology in the modern era. *Gut* **43**: 721–727.
- Blitz J, Smith JW, Gerard JR. 1963. Aloe vera gel in peptic ulcer therapy: preliminary report. *J Am Ost Ass* **62**: 731–735.
- Bronner C, Landry Y. 1985. Kinetics of the inhibitory effect of flavonoids on histamine secretion from mast cells. *Agents Actions* **16**: 147–151.
- Brzozowski T, Konturek SJ, Kwiecin S *et al.* 1998. SU-840, a novel synthetic flavonoid derivative of sophoradin, with potent gastroprotective and ulcer healing activity. *J Physiol Pharmacol* **49**: 83–98.
- Capasso F, Grandolini G. 1999. *Fitofarmacia*. Springer Verlag Italia: Milan.
- Cavallini L, Bindoli A, Siliprandi N. 1978. Comparative evaluation of antiperoxidative action of silymarin and other flavonoids. *Pharmacol Res Commun* **10**: 133.
- Cristoni A, Magistretti MJ. 1987. Antiulcer and healing activity of *Vaccinium myrtillus* anthocyanosides. *II Farmaco* **2**: 29–43.
- De Pasquale R, Germano MP, Keita A, Sanogo R, Iauk L. 1995. Antiulcer activity of *Pteleopsis suberosa*. *J Ethnopharmacol* **47**: 55–58.
- Desai HG, Venugopalan K, Antia FP. 1972. Effect of red chilli powder on DNA content of gastric aspirates. *Gut* **14**: 974–976.
- Di Carlo G, Mascolo N, Izzo AA, Capasso F. 1999. Flavonoids: old and new aspects of a class of natural therapeutic drugs. *Life Sci* **64**: 337–353.
- Di Carlo G, Mascolo N, Izzo AA, Capasso F, Autore G. 1994. Effect of quercetin on the gastrointestinal tract in rats and mice. *Phytother Res* **8**: 42–45.
- Disi AM, Tamimi SO, Abuereish GM. 1998. Effects of *Anchusa strigosa* root aqueous extract on gastric ethanol-induced ulcer in laboratory animals. *J Ethnopharmacol* **60**: 189–198.
- Domschke W, Domschke S, Hagel J, Demling L, Croft DN. 1977. Gastric epithelial cell turnover mucus production, and healing of gastric ulcers with carbenoxolone. *Gut* **18**: 817–820.
- Ezaki N, Kato M, Takizawa N, Morimoto S, Nonaka G, Nishioka I. 1985. Pharmacological studies on *Linderae umbellatae* ramus, IV. Effects of condensed tannin related compounds on peptic activity and stress-induced gastric lesions in mice. *Planta Med* **51**: 34–38.
- Franco L, Manara P, Erbeti I, Velo GP. 1993. Antiulcer activity of carbenoxolone and ISFD 3401 on PGE<sub>2</sub> release in rat gastric mucosa. *Pharmacol Res* **27**: 141–150.
- Galal EE, Kandil A, Hegazy R. 1975. *Aloe vera* and gastrogenic ulceration. *J Drug Res* **7**: 73.
- Gamberini MT, Skorupa LA, Souccar C, Lapa AJ. 1991. Inhibition of gastric secretion by a water extract from *Baccharis triptera*, Mart. *Mem Inst Oswaldo Cruz* **86** (Suppl 2): 137–139.
- Garg GP, Nigam SK, Ogle CW. 1993. The gastric antiulcer effects of the leaves of the *Neem* tree. *Planta Med* **59**: 215–217.
- Germanò MP, De Pasquale R, Iauk L, Galati EM, Keita A, Sanogo R. 1996. Antiulcer activity of *Vernonia Kotschyana* sch. blp. *Phytomedicine* **2**: 229–233.
- Germanò MP, Sanogo R, Guglielmo M, De Pasquale R, Crisafi G, Bisignano G. 1998. Effects of *Pteleopsis suberosa* extracts on experimental gastric ulcer and *Helicobacter pylori* growth. *J Ethnopharmacol* **59**: 167–172.
- Gharzouli K, Khennouf S, Amira S, Gharzouli A. 1999. Effects of aqueous extracts from *Quercus ilex* L. root bark, *Punica granatum* L. fruit peel and *Artemisia herba-alba* Asso leaves on ethanol-induced gastric damage in rats. *Phytother Res* **13**: 42–45.
- Habib A, El-Sebakhy NA, Kadry HA. 1979. New and simple

- methylene blue colorimetric assay for glycyrrhizin in pharmaceuticals. *J Pharm Sci* **68**: 1221–1223.
- Harju E. 1984. Guar gum benefits duodenal ulcer patients by decreasing gastric acidity and rate of emptying of gastric contents 60 to 120 minutes postprandially. *Am Surg* **50**: 668–672.
- Harju E. 1985. Effects of meal viscosity on serum gastrin response and gastric emptying rate in duodenal ulcer patients. *Chir Ital* **37**: 139–147.
- Harju EJ, Larmi TK. 1985. Effect of guar gum added to the diet of patients with duodenal ulcer. *J Parenter Enter Nutr* **9**: 496–500.
- Harju E, Sajanti J. 1987. The protective effect of nutrients against stress induced gastric ulcers in the rat. *Surg Gynecol Obstet* **165**: 530–534.
- Harju E, Sajanti J. 1991. The protective effect of guar gum against stress-induced gastric ulcer in the rat. *In Vivo* **5**: 397–400.
- Hersey SJ. 1994. Gastric secretion of pepsins. In *Physiology of the Gastrointestinal Tract*, Johnson LR (ed.). Raven Press: New York; 1227–1238.
- Hossenbocus A, Colin-Jones DG. 1974. Proceedings: The effects of aspirin, carbenoxolone, and gefarnate on the gastric mucosal potential difference in man. *Gut* **15** (4): 335–336.
- Huong NT, Matsumoto K, Watanabe H. 1998. The antistress effect of majonoside-R2, a major saponin component of Vietnamese ginseng: neuronal mechanisms of action. *Methods Find Exp Clin Pharmacol* **20**: 65–76.
- Iatsynov AI, Belova LF, Lipkina GS, Sokolov SIA, Trutneva EA. 1978. Pharmacology of calendulose B, a new triterpene glycoside from the roots of *Calendula officinalis*. *Farmakol Toksikol* **41**: 556–560.
- Izzo AA, Di Carlo G, Mascolo N, Autore G, Capasso F. 1994. Antiulcer effect of flavonoids. Role of endogenous PAF. *Phytother Res* **6**: 179–181.
- John TA, Onabanjo AO. 1990. Gastroprotective effects of an aqueous extract of *Entandrophragma utile* bark in experimental ethanol-induced peptic ulceration in mice and rats. *J Ethnopharmacol* **29**: 87–93.
- Johnston B, Lindup WE, Shillingford JS, Smith M, Parke DV. 1975. The pharmacological biochemistry of carbenoxolone. Its effects on gastric mucus. In *Fourth Symposium on Carbenoxolone*, Aveny Jones F, Parke DV (eds). Butterworths: London; 3–21.
- Kang JY, Tay HH, Guan R. 1992. Chronic upper abdominal pain: site and radiation in various structural and functional disorders, and the effect of various foods. *Gut* **33**: 743–748.
- Kang JY, Teng CH, Wee A, Chen FC. 1995a. Effect of capsaicin and chilli on ethanol induced gastric mucosal injury in the rat. *Gut* **36**: 664–669.
- Kang JY, Yap I, Guan R, Lim TC. 1988. Chilli ingestion does not lead to macroscopic gastroduodenal mucosa damage in human subjects. *J Gastroenterol Hepatol* **3**: 573–576.
- Kang JY, Yeoh KG, Chia HP *et al.* 1995b. Chilli-protective factor against peptic ulcer? *Dig Dis Sci* **40**: 576–579.
- Katoh Y, Tanaka M, Kawashima H. 1998. Protective effects of teprenone and gefarnate against taurocholate/hydrochloric acid-induced acute gastric mucosal lesions in rats. *Nippon Yakurigaku Zasshi* **112**: 323–331.
- Kent Lloyd KC, Debas HT. 1994. Peripheral regulation of gastric acid secretion. In *Physiology of the Gastrointestinal Tract*, Johnson LR (ed.). Raven Press: New York, 1126–1185.
- Konturek SJ, Radecki T, Brzozowski T *et al.* 1986. Antiulcer and gastroprotective effects of Solon, a synthetic flavonoid derivative of sophoradin. Role of endogenous prostaglandins. *Eur J Pharmacol* **125**: 185–192.
- Koo MW. 1994. *Aloe vera*: antiulcer and antidiabetic effects. *Phytother Res* **8**: 461–464.
- Kulkarni SK, Goel RK. 1996. Gastric antiulcer activity of UL-409 in rats. *Indian J Exp Biol* **34**: 683–688.
- Kumar N, Vij JC, Sarin SK, Anand BS. 1984. Do chillies influence the healing of duodenal ulcer? *Br Med J* **288**: 1803–1804.
- Kyogoku K, Katayama K, Yokomori S *et al.* 1979. Anti-ulcer effect of isoprenyl flavonoids. Synthesis and antiulcer activity of new chalone related to sophoradin. *Chem Pharm Bull* **27**: 2943–2948.
- Lambrecht N, Burchert M, Respondek M, Muller KM, Peskar BM. 1993. Role of calcitonin gene-related peptide and nitric oxide in the gastroprotective effect of capsaicin in the rat. *Gastroenterology* **104**: 1371–1380.
- Lewis DA, Fields WN, Shaw GP. 1999. A natural flavonoid present in unripe plantain banana pulp (*Musa sapientum* L. var. *paradisica*) protects the gastric mucosa from aspirin-induced erosions. *J Ethnopharmacol* **65**: 283–288.
- Lorenz W, Reimann HJ, Kusche J *et al.* 1975. Effects of catechin on several enzymes of histamine metabolism and on stress ulcer formation in the female rat. *Naunyn Schmiedebergs Arch Pharmacol* **287** Suppl: R62.
- Magistretti MJ, Conti M, Cristoni A. 1988. Antiulcer activity of an anthocyanidin from *Vaccinium myrtillus*. *Arzneim Forsch* **38**: 686–690.
- Maity S, Vedasiromoni JR, Ganguly DK. 1995. Anti-ulcer effect of the hot water extract of black tea (*Camellia sinensis*). *J Ethnopharmacol* **46**: 167–174.
- Mandal S, Das DN, De K *et al.* 1993. *Ocimum sanctum* Linn. A study on gastric ulceration and gastric secretion in rats. *Indian J Physiol Pharmacol* **37**: 91–92.
- Manicheva OA, Barnaulov OD. 1984. Effect of plant drugs on the nucleic acid content of gastric tissues during ulcerogenesis. *Rastit Resu* **20**: 256–264.
- Manonmani S, William S, Subramanian S, Govindasamy S. 1994. Biochemical evaluation of the antiulcerogenic effect of Cauvery-100 (an ayurvedic formulation) in rats. *J Ethnopharmacol* **42**: 1–5.
- Marhuenda E, Alarcon de la Lastra C, Martin MJ. 1994. Antisecretory and gastroprotective effects of aescine in rats. *Gen Pharmacol* **25**: 1213–1219.
- Marhuenda E, Martin MJ, Alarcon de la Lastra C. 1993. Antiulcerogenic activity of aescine in different experimental models. *Phytother Res* **7**: 13–16.
- Mariko M. 1989. *Etude de l'activité du 'terenifou', écorce du tron de Pteleopsis suberosa Engl. Et Diels (Combretaceae) dans le traitement des ulcères gastro-duodénaux. Thèse de Médecine*, Bamako, Mali.
- Marti-Bonmati E, Alino SF, Llorisy SM, Esplugues J. 1980. Effects of cimetidine, atropine and prostaglandin E<sub>2</sub> on rat mucosal erosions produced by intragastric distension. *Eur J Pharmacol* **68**: 49–53.
- Martin MJ, Alarcon de la Lastra C, Marhuenda E, Delgado F. 1988. Anti-ulcerogenicity of the flavonoid fraction from *Ditrichia viscosa* (L.) W. Greuter, in rats. *Phytother Res* **2**: 183–186.
- Martin MJ, La Casa C, Alarcon de la Lastra C, Cabeza J, Villegas I, Motilva V. 1998. Anti-oxidant mechanisms involved in gastroprotective effects of quercetin. *Z Naturforsch* **53**: 82–88.
- Martin MJ, Marhuenda E, Perz-Guerrero C, Franco JM. 1994. Antiulcer effect of naringin on gastric lesions induced by ethanol in rats. *Pharmacology* **49**: 144–150.
- Martin MJ, Motilva V, Alarcon de la Lastra C. 1993. Quercetin and naringenin: Effects on ulcer formation and gastric secretion in rats. *Phytother Res* **7**: 150–153.
- Matsuda H, Li Y, Murakami T, Yamahara J, Yoshikawa M. 1998. Protective effects of oleanolic acid oligoglycosides on ethanol- or indomethacin-induced gastric mucosal lesions in rats. *Life Sci* **63**: PL245–250.
- Mitra SK, Gopumadhavan S, Hemavathi TS, Muralidhar TS, Venkataranganna MV. 1996. Protective effect of UL-409, a herbal formulation against physical and chemical factor induced gastric and duodenal ulcers in experimental animals. *J Ethnopharmacol* **52**: 165–169.
- Morgan RJ, Nelson LM, Russell RI, Docherty C. 1983. The protective effect of deglycyrrhized liquorice against aspirin and aspirin plus bile acid-induced gastric mucosal damage, and its influence on aspirin absorption in rats. *J Pharm Pharmacol* **35**: 605–607.
- Moroney MA, Alcaraz MJ, Folder RA, Carey F, Hoult SRS. 1988. Selectivity of neutrophil 5-lipoxygenase and cyclooxygenase inhibition by anti-inflammatory flavonoid glycoside and related aglycone flavonoids. *J Pharm Pharmacol* **40**: 787–792.
- Motilva V, Alarcon De La Lastra C, Martin MJ. 1992. Effects of naringenin and quercetin on experimental chronic gastric ulcer in rats: Studies on the histological findings. *Phytother Res* **6**: 168–170.
- Motilva V, Alarcon de la Lastra C, Martin MJ. 1993. Ulcer

- protecting effects of naringenin on gastric lesions induced by ethanol in rat: Role of endogenous prostaglandins. *J Pharm Pharmacol* **46**: 91–94.
- Myers BM, Lacey Smith J, Graham DY. 1987. Effect of red pepper and black pepper on the stomach. *Am J Gastroenterol* **82**: 211–214.
- Navarrete A, Martinez-Urbe LS, Reyes B. 1998. Gastroprotective activity of the stem bark of *Amphipterygium adstringens* in rats. *Phytother Res* **12**: 1–4.
- Newall CA, Anderson LA, Phillipson JD. 1996. *Herbal Medicines*. The Pharmaceutical Press: London.
- Nguyen TT, Matsumoto K, Yamasaki K, Nguyen MD, Nguyen TN, Watanabe H. 1996. Effects of majonoside-R2 on pentobarbital sleep and gastric lesion in psychologically stressed mice. *Pharmacol Biochem Behav* **53**: 957–963.
- Ottenjann R, Rosch W. 1970. Therapie des Ulcus ventriculi mit Carbenoxolon-Natrium. Ergebnisse eines Doppelblindversuchs. *Med Klin* **65**: 74–80.
- Pal S, Nag Chaudhuri AK. 1991. Studies on the anti-ulcer activity of a *Bryophyllum pinnatum* leaf extract in experimental animals. *J Ethnopharmacol* **33**: 97–102.
- Parmar NS. 1983. The gastric antiulcer activity of naringenin, a specific histidine decarboxylase inhibitor. *Int J Tiss Res* **4**: 415–420.
- Parmar NS, Tariq M, Al-Yahya MA, Ageel AM, Al-Said MS. 1986. Evaluation of *Aloe vera* leaf exudate and gel for gastric and duodenal anti-ulcer activity. *Fitoterapia* **57**: 380–383.
- Peskar BM. 1980. Effect of carbenoxolone on prostaglandin synthesizing and metabolizing enzyme and correlation with gastric mucosal carbenoxolone concentrations. *Scand J Gastroenterol* **15** (Suppl 65): 109–112.
- Peskar BM, Respondek M, Muller KM, Peskar BA. 1991. A role for nitric oxide in capsaicin-induced gastroprotection. *Eur J Pharmacol* **198**: 113–114.
- Rafatullah S, Tariq M, Al-Yahya MA, Mossa JS, Ageel AM. 1990. Evaluation of turmeric (*Curcuma longa*) for gastric and duodenal antiulcer activity in rats. *J Ethnopharmacol* **29**: 25–34.
- Rafatullah S, Tariq M, Mossa JS, Al-Yahya MA, Al-Said MS, Ageel AM. 1993. Protective effect of *Swertia chirata* against indomethacin and other ulcerogenic agent-induced-gastric ulcers. *Drugs Exp Clin Res* **19**: 69–73.
- Rainova L, Nakov N, Bogdanova S, Minkov E, Staneva-Stoytcheva D. 1988. Ulceroprotective activity of the flavonoids of *Genista rumelica* Vel. *Phytother Res* **2**: 137–139.
- Rao VS, Santos FA, Sobreira TT, Souza MF, Melo CL, Silveira ER. 1997. Investigations on the gastroprotective and antidiarrhoeal properties of ternatin, a tetramethoxyflavone from *Egletes viscosa*. *Planta Med* **63**: 146–149.
- Rao VS, Viana GS, Menenez AM, Gadelha MG. 1987. Studies on the antiulcerogenic activity of *Astronium urundeuva* Engl. II. Aqueous extract. *Braz J Med Biol Res* **20**: 803–805.
- Rees WDW, Rhodes J, Wright JE, Stamford IF, Bennett A. 1979. Effect of deglycyrrhized liquorice on gastric mucosal damage by aspirin. *Scand J Gastroent* **14**: 605–607.
- Reyes M, Martin C, Alarcon de la Lastra C, Trujillo J, Toro MV, Ayuso MJ. 1996. Antiulcerogenicity of the flavonoid fraction from *Erica andevalensis* Cabezudo-Rivera. *Z Naturforsch* **51**: 563–569.
- Reynolds T, Dweck AC. 1999. *Aloe vera* leaf gel: a review update. *J Ethnopharmacol* **68**: 3–37.
- Robak J, Gryglewski R. 1988. Flavonoids are scavengers of superoxide anions. *Biochem Pharmacol* **37**: 837–841.
- Robak J, Shridi F, Wolbis M, Kerolikowska M. 1988. Screening of the influence of flavonoids on lipoxygenase and cyclooxygenase activity in lipid oxidation. *Pol J Pharmacol* **40**: 451–458.
- Saijo R, Nonana G, Nishioka I. 1989. Tannins and related compounds. LXXXIV. Isolation and characterization of five new hydrolyzable tannins from the bark of *Mallotus japonicus*. *Chem Pharm Bull* **37**: 2063–2070.
- Saito H, Imanishi K, Okabe S. 1989. Effects of aloe extract, Aloctin A on gastric secretion and on experimental gastric lesions in rats. *Yakugaku Zasshi* **109**: 335–339.
- Saito H, Lee YM, Takagi K, Shoji S, Kondo N. 1977. Pharmacological studies of *Panacis japonici* rhizoma. I. *Chem Pharm Bull* **25**: 1017–1025.
- Saitoh T, Kinoshita T, Shibata S. 1976. Flavonols of licorice root. *Chem Pharm Bull (Tokyo)* **24**: 1242–1245.
- Salvayre R, Braquet P, Perruchot L, Douste-Blazy L. 1982. Comparison of the scavenger effect of bilberry anthocyanosides with various flavonoids. *Flavonoids Bioflavonoids* **11**: 437–442.
- Samuelsson G. 1999. *Drugs of Natural Origin*. Swedish Pharmaceutical Press: Sweden.
- Sanogo R, De Pasquale R, Germanò MO, Iauk L, De Tommasi N. 1996. *Vernonia kotschyana* Sch. Bip.: tolerability and gastroprotective activity. *Phytother Res* **10**: S169–S171.
- Scarlat M, Sandor V, Tamas M, Cuparencu B. 1985. Experimental anti-ulcer activity of *Veronica officinalis* L. extracts. *J Ethnopharmacol* **13**: 157–163.
- Sen T, Pal S, Nag Chaudhuri AK. 1992. Studies on the anti-ulcer activities of *Pluchea indica* less. In *Natural Drugs and the Digestive Tract*, Capasso F, Mascolo N (eds). EMSI: Rome; 69–72.
- Singh S, Majumdar DK. 1999. Evaluation of the gastric antiulcer activity of fixed oil of *Ocimum sanctum* (Holy Basil). *J Ethnopharmacol* **65**: 13–19.
- Souza-Formigoni ML, Oliveira MG, Monteiro MG, da Silveira-Filho NG, Braz S, Carlini EA. 1991. Antiulcerogenic effects of two *Maytenus* species in laboratory animals. *J Ethnopharmacol* **34**: 21–27.
- Suffredini IB, Bacchi EM, Sertie JA. 1999. Antiulcer action of *Microgramma squamulosa* (Kaulf.) Sota. *J Ethnopharmacol* **65**: 217–223.
- Szallasi A, Blumberg PM. 1999. Vanilloid (capsaicin) receptors and mechanisms. *Pharmacol Rev* **51**: 159–221.
- Takagi K, Ishii, Y. 1967. Peptic ulcer inhibiting properties of a new fraction from licorice root (FM100). I. Experimental peptic ulcer and general pharmacology. *Arzneim Forsch* **17**: 1544–1548.
- Takagi K, Okabe S, Saziki R. 1969. A new method for the production of chronic gastric ulcer in rats and the effect of several drugs on its healing. *Japan J Pharmacol* **19**: 418–422.
- Takano F, Yoshizaki F, Suzuki K, Suya N, Ando T, Hisamichi S. 1990. Anti-ulcer effects of *Trichosanthes* fruits. *Chem Pharm Bull (Tokyo)* **38**: 1313–1316.
- Tan PV, Nditafon NG, Yewah MP, Dimo T, Ayafor FJ. 1996. *Eremomastax speciosa*: effects of leaf aqueous extract on ulcer formation and gastric secretion in rats. *J Ethnopharmacol* **54**: 139–142.
- Teng CH, Kang JY, Wee A, Lee KO. 1998. Protective action of capsaicin and chilli on haemorrhagic shock-induced gastric mucosal injury in the rat. *J Gastroenterol Hepatol* **13**: 1007–1014.
- Tepperman BL, Jacobson ED. 1994. Circulatory factors in gastric mucosal defense and repair. In *Physiology of the Gastrointestinal Tract*, Johnson LR (ed.). Raven Press: New York; 1331–1352.
- Teradaira R, Shinzato M, Beppu H, Fujita K. 1993. Anti-gastric ulcer effects of *Aloe arborescens* Mill. Var *natalensis* Berber. *Phytother Res* **7**: S34–S36.
- Tournier H, Schinella G, de Balsa EM, Buschiazzi H, Manez S, Mordujovich de Buschiazzi P. 1999. Effect of the chloroform extract of *Tanacetum vulgare* and one of its active principles, parthenolide, on experimental gastric ulcer in rats. *J Pharm Pharmacol* **51**: 215–219.
- Van Huis GA, Kramer MF. 1981. Effects of carbenoxolone on the synthesis of glycoproteins and DNA in rat gastric epithelial cells. *Gut* **22**: 782–787.
- Vela SM, Souccar C, Lima-Landman MT, Lapa AJ. 1997. Inhibition of gastric acid secretion by the aqueous extract and purified extracts of *Stachytarpheta cayennensis*. *Planta Med* **63**: 36–39.
- Villar A, Gasco MA, Alcaraz MJ. 1984. Anti-inflammatory and anti-ulcer properties of hypolaetin-8-glucoside, a novel plant flavonoid. *J Pharm Pharmacol* **36**: 820–823.
- Yamahara J, Kobayashi M, Miki K, Kuzoka M, Sawada T, Fujimura H. 1985. Cholagogic and antiulcer effect of *Saussurea* radix and its active components. *Chem Pharm Bull* **33**: 1285–1288.
- Yamahara J, Kubomura Y, Miki K, Fujimura H. 1987. Anti-ulcer action of *Panax japonicus* rhizome. *J Ethnopharmacol* **19**: 95–101.

- Yamahara J, Mochizuki M, Fujimura H *et al.* 1990. Antiulcer action of *Sophora flavescens* root and an active constituent. I. *J Ethnopharmacol* **29**: 173–177.
- Yamahara J, Mochizuki M, Rong HQ, Matsuda H, Fujimura H. 1988. The anti-ulcer effect in rats of ginger constituents. *J Ethnopharmacol* **23**: 299–304.
- Yamamoto I. 1973. Aloe ulcin, a new principle of Cape aloe and gastrointestinal function, especially experimental ulcer in rats. *J Med Soc Toho University* **20**: 342–347.
- Yeoh KG, Kang JY, Yap I *et al.* 1995. Chilli protects against aspirin-induced gastroduodenal mucosal injury in humans. *Dig Dis Sci* **40**: 480–583.
- Yesilada E, Takaishi Y. 1999. A saponin with anti-ulcerogenic effect from the flowers of *Spartium junceum*. *Phytochemistry* **51**: 903–908.